Effect of *Cymbopogon citratus* Stapf.
on renal functions in dogs.


Crude water extracts of *Cymbopogon citratus* (*C. citratus*) 1.25, 2.5, 5 and 10 g/kg-bw. was given orally via oro-gastric tubes in dogs anesthetized with sodium pentobarbital. Renal functions were elucidated every 30 min in the 4 hours following administration. The results found insignificant changes of circulatory hemodynamics and renal functions in 1.25, 2.5 and 5 g/kg-bw. dogs. Dogs receiving 10 g/kg-bw *C. citratus* showed significant decreases in heart rate during the first 1.5 – 2.5 hours. Urine volume, glomerular filtration rate and renal plasma flow decreased, while hematocrit increased significantly with slight change of the filtration fraction. Significant decreases in the plasma clearance of osmolality, accompanied with small decreases in free water clearance were demonstrated. Urinary excretion of sodium, potassium and chloride fell significantly with a decrease in fractional excretion. The results could be concluded that oral administration of crude water extracts of *C. citratus* do not have diuretic effects. The increases in urine flow may be due to the drinking of large amounts of water. Furthermore, higher doses may have some toxic effects. However, the mechanism is still unclear and needs to be investigated.

**Key words**: Lemongrass, *Cymbopogon citratus*, Renal functions, Dogs.

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พร้อมสุข ชุตากา, บังอร ชมเขต. ผลของน้ำสกัดตะไคร้ต่อการทำหน้าที่ของไตสุนัข.
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ให้น้ำสกัดตะไคร้วันละ 1, 2.5, 5 และ 10 กรัมต่อน้ำหนักตัว 1 กิโลกรัม ทางปากโดย
ผ่านท่อเข้าสู่กระเพาะอาหารก่อนสุนัขที่ตอบโต้โดยโจมตีพบไม่มีการปิดลิด ตรวจวัดการทำงานของ
ไตของโดยทุกครั้งวันละจนครบ 4 ชั่วโมงหลังให้น้ำสกัดตะไคร้. ผลการทดลองพบว่าน้ำสกัดตะไคร้
วันละ 1, 2.5, 5 และ 10 กรัมต่อน้ำหนักตัว 1 กิโลกรัมไม่มีเปลี่ยนแปลงพลวัตของเลือดและการทำ
หน้าที่ของไต สุนัขกลุ่มที่ได้รับน้ำสกัดตะไคร้วันละ 10 กรัมต่อน้ำหนักตัว 1 กิโลกรัมนั้นพบว่า
อัตราการปั้มวันละทำไต[length]นโยบายมีน้อยกว่าน้ำสกัดตะไคร้ในช่วงเวลา 1.5-2.5 ชั่วโมง การทดสอบ
ปัจจุบัน อัตราการปั้ม และปริมาตรพัสตราขณะใดตลอดขณะที่ปริมาตรพัสตราเม็ดเลือดเพิ่มขึ้น
อย่างมีนัยสำคัญทางสถิติ พร้อมกับมีลดส่วนของอัตราการปั้มตลอดเวลาในผลิตภัณฑ์พบเพียงเล็กน้อย
การส่งออกผลิตภัณฑ์สูงกว่าในผลิตภัณฑ์โดยตลอดและที่ให้กับเจ้าหน้าที่สาระได้เพียงเล็กน้อย
อัตราการชี้ขาดยกโดยไม่ได้ ไปแสดงชัดเจน และแสดงว่าการทดลองผลิตภัณฑ์มีนัยสำคัญทางสถิติ
พร้อมกับสอดส่วนของการขับออกต่ออัตราการปั้มน้ำ ผลของการศึกษาครั้งนี้จะสรุปได้ว่า
การดื่มน้ำสกัดตะไคร้ไม่มีผลอัตราการขับปัสสาวะ ภูมิคุ้มกันที่มีต่อการ
ขับปัสสาวะจะเกิดจากการดื่มน้ำสกัดตะไคร้นั้นน้อย อีกว่าการเกิดผลขึ้นนี้น้ำสกัดตะไคร้ที่ขับ
ไตในป่าจะเป็นพิษต่อร่างกายได้ อย่างไรก็ตามพบว่าไม่น้อยดังที่การศึกษาต่อไปอีก
Cymbopogon citratus (lemon grass) is widely employed as a folk medicinal plant for the treatment of several diseases and symptoms. A herbal tea or decoction prepared from the dried leaves is called 'abafado' in Brazil. It is frequently used as a sedative and hypnotic, analgesic, antiemetic, antispasmodic and for other stomach disorders.\(^{(1,2)}\) In Nigeria it is used as an antipyretic and antispasmodic.\(^{(3)}\)

In Angola and India it is considered to be an antitussive, antiemetic, antiseptic and anti-rheumatic agents.\(^{(4)}\) In Indonesia it is employed to help digestion and as a diuretic.\(^{(5)}\) In ancient Thai medicinal treatments, it has been used as carminative, diuretic, antihypertensive, antianorexic, antispasmodic and analgesic agents, and it has been used for urinary tract problems.

Locksley et al. (1982) also reported that hot water extracts of dried leaves and stems of lemongrass has been used as an effective renal antispasmodic and diuretic in Egypt.\(^{(6)}\) However, its action on renal functions has never previously been studied. Our research aimed to study the effects of crude water extracts on renal functions in order to confirm the diuretic effect which was previously conceptualized in many countries, including Thailand. Lemongrass is easily obtained and is very cheap in Thailand. This plant may be used as a medicine in the future.

**Materials and methods**

**Preparation of crude extract of C. citratus**

100 grams of lower lemongrass leaves of which had been dried in an oven at 60-70°C for 24-36 hours then boiled in distilled water for 5 minutes and then readjusted to a volume of 100 ml after filtration through muslin. The decoctions came to 100g/dl.

**Animals and treatment**

Twenty adult male mongrel dogs weighing 12-18 kg were maintained with free access to water and food. Each had been kept for at least 7 days prior to the experiment. They were fasted for 12 hours preceding each operation. Each animal was anesthetized by intravenous injection of 25 mg/kg of sodium pentobarbital. A tracheostomy was performed and the animals were allowed to ventilate spontaneously in room air. Both femoral veins were catheterized, one for infusion of inulin and the other for infusion of normal saline and supplemental doses of sodium pentobarbital.

A catheter was inserted into the right femoral artery and connected to a pressure transducer for recording the systemic arterial blood pressure and heart rate on a Harvard universal oscillograph. The left kidney was exposed retroperitoneal through a flank incision and the left ureter was cunuated. The left renal artery was encircled with an electromagnetic flow probe and connected to electromagnetic blood flowmeter for recording the renal blood flow.

During the surgical and experimental procedures, isotonic saline was infused continuously at a rate 1.0-1.5 ml/min in order to maintain the extracellular fluid volume. A priming dose of 50 mg/kg inulin was administered, followed by sustaining infusion sufficient to maintain the plasma inulin concentration at approximately 20 mg/dl. A period of 50 min was allowed to elapse for stabilization of the general condition and plasma inulin concentration. Blood and urine samples were obtained for two control periods of 30 minutes. Arterial blood was drawn at the midpoint of each urine collection period.
C. citratus in 10 ml doses of 1.25, 2.5, 5 or 10g/kg was intragastrically administered as a single dose in each group of 5 dogs. The concentrations of sodium and potassium, chloride and inulin were determined by use of a flame photometer (KLiNa Flame Operating–Bechman Instrument, Instrument Lab. model 343), chloride analyzer (Bechman Instrument), and Jaffy reaction, respectively. Plasma osmolality was measured by freezing point depression technique. Hematocrit was also determined from each blood sample.

At the end of experiment the left kidney was excised, stripped of surrounding fat and tissue, blotted dry and weighed so that the kidney functions could be expressed as per gram kidney weight. The results are shown as mean ± SEM between before and after feeding. The statistical significance was assessed by using the Student’s paired t-test with a p value less than .05.

**Results**

**Effect of C. citratus on mean arterial pressure, heart rate and hematocrit.**

Figure 1 demonstrated that during experimental period, there were slightly changes in mean arterial pressure (MAP), heart rate (HR) and hematocrit (Hct) in small doses until 5 g/kg. In 10 g/kg dogs, the increase of MAP and the decrease of HR were significant during 1-2.5 hrs as compared to the control value. Although the Hct was significantly increased throughout the experimental period as shown in figure 1 and table 1.

**Effect of C. citratus on urine flow rate, renal plasma flow, glomerular filtration rate and filtration fraction.**

As shown in figure 2 and table 1, the urine flow rate (V) was significantly decreased in 10 g/kg

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**Figure 1.** Effect of C. citratus 1.25, 2.5, 5 and 10 g/kg on mean ± SEM of mean arterial pressure (MAP), heart rate (HR) and hematocrit (Hct).

* *p < .05, **p < .01, ***p < .005

**Figure 2.** Effect of C. citratus 1.25, 2.5, 5 and 10 g/kg on mean ± SEM of urine flow rate (V), renal plasma flow (RPF), glomerular filtration rate (GFR) and filtration fraction (FF).

* *p < .05, **p < .01, ***p < .005
Table 1. The significant changes in mean ± SEM of renal and circulatory hemodynamics following C. citratus administration.

<table>
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<tr>
<th>Parameters</th>
<th>Doses g/kg</th>
<th>0</th>
<th>0.5</th>
<th>1.0</th>
<th>1.5</th>
<th>2.0</th>
<th>2.5</th>
<th>3.0</th>
<th>3.5</th>
<th>4.0</th>
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<tr>
<td>HR beat/min</td>
<td>10</td>
<td>122.20</td>
<td>120.00</td>
<td>110.80</td>
<td>109.60**</td>
<td>103.4***</td>
<td>110.40*</td>
<td>119.00</td>
<td>127.20</td>
<td>123.00</td>
</tr>
<tr>
<td>Hct %</td>
<td>10</td>
<td>29.40</td>
<td>31.20*</td>
<td>32.60*</td>
<td>33.40**</td>
<td>32.40*</td>
<td>35.00*</td>
<td>34.80*</td>
<td>34.80*</td>
<td>37.00</td>
</tr>
<tr>
<td>V μl/min-g kw</td>
<td>10</td>
<td>12.48</td>
<td>4.59*</td>
<td>3.46*</td>
<td>3.26*</td>
<td>3.34*</td>
<td>3.73</td>
<td>3.46</td>
<td>3.95</td>
<td>5.75</td>
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<tr>
<td>RPF ml/min-g kw</td>
<td>1.25</td>
<td>2.47</td>
<td>2.08*</td>
<td>2.17</td>
<td>2.14</td>
<td>2.11</td>
<td>2.12</td>
<td>2.08</td>
<td>2.20</td>
<td>2.21</td>
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<tr>
<td>RPF ml/min-g kw</td>
<td>2.5</td>
<td>2.86</td>
<td>2.12*</td>
<td>1.99*</td>
<td>2.09*</td>
<td>2.15*</td>
<td>2.33</td>
<td>2.33</td>
<td>2.52</td>
<td>2.50</td>
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<td>RPF ml/min-g kw</td>
<td>5</td>
<td>4.74</td>
<td>4.09*</td>
<td>3.86*</td>
<td>4.10</td>
<td>4.04</td>
<td>4.29</td>
<td>4.39</td>
<td>4.44</td>
<td>4.57</td>
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<tr>
<td>RPF ml/min-g kw</td>
<td>10</td>
<td>2.99</td>
<td>2.13***</td>
<td>1.87***</td>
<td>1.83***</td>
<td>1.98**</td>
<td>1.94***</td>
<td>1.86*</td>
<td>1.86***</td>
<td>1.85**</td>
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<td>GFR ml/min-g kw</td>
<td>0.47</td>
<td>0.22*</td>
<td>0.29*</td>
<td>0.31</td>
<td>0.42</td>
<td>0.42</td>
<td>0.30</td>
<td>0.33</td>
<td>0.35</td>
<td></td>
</tr>
</tbody>
</table>

* P<0.05, ** P<0.01, *** P<0.005

kg dogs at the period of 2 hrs. The decrease of renal plasma flow (RPF) was seen significantly in 1.25 g/kg dogs at the period of 30 min. However, this decrease of RPF was stayed for 2 hrs in 2.5 g/kg dogs. In addition, the results were shown that RPF of the 5 g/kg dogs was significantly decreased during the first hour as compared to the control value.

At the highest dose (10 g/kg), the RPF was decreased significantly throughout the experimental period. Besides, during the first hour, the glomerular filtration rate (GFR) was decreased significantly without significant change in filtration fraction (FF) throughout the experimental period as demonstrated in figure 2.
Effect of *C. citratus* on plasma concentration, excretion rate and fractional excretion of sodium.

The results shown in figure 3 and table 2 indicated that plasma concentration of sodium (P<sub>Na</sub>) was not altered after administration of *C. citratus*, while there were significant decreases of excretion rate (U<sub>Na</sub>V) and fractional excretion (FENa) except dogs that received 2.5 g/kg of *C. citratus*. The reduction of U<sub>Na</sub>V in 1.25 g/kg dogs was observed during 1.5 hrs. However, at 5 g/kg, U<sub>Na</sub>V and FENa were significantly decreased observed only at one hour period after the *C. citratus* administration. The results monitored from 10 g/kg indicated that U<sub>Na</sub>V was significantly decreased during the period of 0.5-3.5 hrs. In addition, FENa was significantly decreased during the period of 1.0-2.5 hrs as shown in table 2.

![Figure 3](image)

**Figure 3.** Effect of *C. citratus* 1.25, 2.5, 5 and 10 g/kg on mean ± SEM of plasma concentration of sodium (P<sub>Na</sub>), excretion rate of sodium (U<sub>Na</sub>V) and fractional excretion of sodium (FENa).

*<sup>p</sup> < .05, **<sup>p</sup> < .01, ***<sup>p</sup> < .005

Effect of *C. citratus* on plasma concentration, excretion rate and fractional excretion of potassium.

The results shown in figure 4 and table 2 indicated that the plasma concentration of potassium (P<sub>K</sub>) was significantly increased during 1-2.5 hrs, while the excretion rate of potassium (U<sub>K</sub>V) was significantly decreased during the period of 0.5-1.5 hrs after the administration of 10 g/kg *C. citratus* without significantly changes of fractional excretion of potassium (FENa).

![Figure 4](image)

**Figure 4.** Effect of *C. citratus* 1.25, 2.5, 5 and 10 g/kg on mean ± SEM of plasma concentration of potassium (P<sub>K</sub>), excretion rate of potassium (U<sub>K</sub>V) and fractional excretion of potassium (FENa).

*p < .05, **p < .01, ***p < .005

Effect of *C. citratus* on plasma concentration, excretion rate and fractional excretion of chloride.

There were slight changes in plasma concentration of chloride (P<sub>Cl</sub>) as demonstrated in figure 5. The significant decrease in excretion rate (U<sub>Cl</sub>V) during 1 to 3 hrs after the administration of 10 g/kg was observed. However, the reduction of fractional excretion (FENa) was only observed at 2 hrs as shown in figure 5 and table 2.
Table 2. The significant changes in mean ± SEM of electrolytes and osmolality following *C. citratus* administration.

<table>
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<th>Parameters</th>
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<td></td>
<td>g/kg</td>
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<tr>
<td></td>
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</tr>
<tr>
<td></td>
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<td>±0.33</td>
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<tr>
<td>UNaV µEq/min-g kw</td>
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<td>1.75</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>1.33</td>
</tr>
<tr>
<td></td>
<td></td>
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<tr>
<td>FE Na%</td>
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</tr>
<tr>
<td></td>
<td>10</td>
<td>1.99</td>
</tr>
<tr>
<td></td>
<td></td>
<td>±0.28</td>
</tr>
<tr>
<td></td>
<td></td>
<td>±0.16</td>
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<td>UKV µEq/min-g kw</td>
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</tr>
<tr>
<td></td>
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<tr>
<td>UClV µEq/min-g kw</td>
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</tr>
<tr>
<td></td>
<td></td>
<td>±0.33</td>
</tr>
<tr>
<td>FE Cl%</td>
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</tr>
<tr>
<td></td>
<td></td>
<td>±0.62</td>
</tr>
<tr>
<td>C Osm µl/min-g kw</td>
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<td>14.96</td>
</tr>
<tr>
<td></td>
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<td>±3.24</td>
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</table>

*p<.05, **p<.01, ***<.005*
Figure 5. Effect of *C. citratus* 1.25, 2.5, 5 and 10 g/kg on mean ± SEM of plasma concentration of chloride (P_{Cl}), excretion rate of chloride (U_{ClV}) and fractional excretion of chloride (FECI).

* *p < .05, **p < .01, ***p < .005

Figure 6. Effect of *C. citratus* 1.25, 2.5, 5 and 10 g/kg on mean ± SEM of plasma osmolality (P_{Osm}), clearance of osmolality (C_{Osm}) and free water clearance (C_{H2O}).

* *p < .05, **p < .01, ***p < .005

Effect of *C. Citratus* on plasma osmolality, plasma clearance of osmolality and free water clearance.

Plasma osmolality (P_{Osm}), plasma clearance of osmolality (C_{Osm}) and free water clearance (C_{H2O}) were altered insignificantly in dogs received 1.25, 2.5 and 5 g/kg as indicated in figure 6. In 10 g/kg, the C_{Osm} was decreased significantly during the period of 0.5–3 hrs without significant changes in C_{H2O} as shown in figure 6 and table 2.

Discussion

Preliminary studies demonstrated that a 0.3 g/kg decoction of *C. citratus* showed hypotensive effects when given intravenously to rats, while 5 g/kg decoction showed diuretic effects when given orally. Conversely, our investigations found that 1.25 and 2.5 g/kg of decoction had no effects on arterial blood pressure whereas 10 g/kg showed slight increases at the period of 1.5–2.5 hrs after feeding and this was accompanied with a significant decrease in HR (figure 1 and table 1).

The increment of vascular resistance may be responsible for the increase in arterial blood pressure that in turn elicited reflex decreases in heart rate and cardiac output via the arterial baroreceptor. In addition, the augmentation of arterial blood pressure is probably due to an increase in blood viscosity which was shown by the hematocrit. The increase in hematocrit was probably caused by an increase in circulating red cell mass induced by splenic contraction as described by Bell et al., (1981). However, this may be unlikely because in the third hour of highest concentration of decoction the MAP returned to its baseline value whereas the Hct continued to increase (figure 1 and table 1).

GFR and RPF were decreased after intragastric administration of 10 g/kg decoction (figure 2 and table 1). These changes may be due to the action of some substances in *C. citratus* on
vascular smooth muscle. The increase in vascular resistance accompanied with hematocrit becomes important in the distal glomerular capillaries and efferent arterioles. In efferent arteriole, Hct was even higher than in the afferent arterioles due to glomerular filtration. Thus, the decrease in GFR was less than RPF resulting increases in FF (figure 2 and table 1).

In addition, the alteration of resistance in renal vessels that decrease GFR and RPF may be caused by the secretion of renin from JG-cells resulting in vasoconstriction. Nevertheless, Linas et al. (1980) reported that pentobarbital anesthesia decreased RBF, increased systemic and renal vascular resistances and contributed to the failure to excrete a normal saline load. (11)

The urine flow rate and osmolar clearance were significantly decreased when the dogs were given the crude extract from C. citratus at 10 g/kg oral doses (figures 2 and 6, tables 1 and 2). These changes may result from the decrease in RPF and GFR. Since vasopressin causes increases of water reabsorption in distal and collecting tubules, the decrease in the urine flow rate and negative free water clearance which was exhibited probably due to the same reason. It has been demonstrated by Robertson (1977) and Goetz et al. (1988) that vomit is a potent stimulus for vasopressin secretion. (9, 12) In the present study, two dogs at the highest dose vomited after receiving the decoction. Therefore, this may suggest that dogs have some increases in circulating vasopressin levels.

The decreases in the urinary excretion rates of sodium, potassium and chloride in 10 g/kg dogs as demonstrated in figures 3, 4, 5 and table 2 may be due to a vasoconstrictive action of some substances in lemongrass causing a substantial decreases in GFR that in turn is responsible for the reduced electrolyte excretion. The significant increases in plasma potassium (P_K) may result from permeability changes in the membrane of erythrocytes or damage of the Na^+, K^+ -pump at this membrane. It would cause leakage of K^+ efflux to blood circulation. However, the hemolysis may not support the change of P_K in this experiment because there was no evidence of hemolysis in the plasma samples.

The increment of P_K is probably due to pseudohyperkalemia while the elevated potassium concentration occurs in vitro not in vivo. (13, 14) In addition, Ifudu et al. (1992) described that a circulating uremic toxin which inhibits cell membrane ouabain sensitive Na^+, K^+ -ATPase has been implicated as the cause of lowered intracellular potassium. (15) They suggested that if this inhibition is present in vitro, it might lead to increased leakage of potassium from cells in serum samples with resultant exaggeration of pseudohyperkalemia in the uremic patients.

The loss of potassium from the leucocytes may increase extracellular potassium concentrations. In undialysed patients with advanced renal failure, leucocyte sodium and water contents were significantly greater than normal, while leucocyte potassium content was reduced. (16) These changes may occur because when potassium is lost from the cells, its replacement by sodium might increase the total osmotically active cation and hence cell water. The alternation may cause the increased plasma potassium and the reduction of sodium excretion in the urine for stability of plasma sodium.

This study could be summarized that the effects of intragastric administration of crude water decoction from C. citratus at concen-
trations of less than 5 g/kg on hemodynamics and renal functions were not different from the baseline control at the starting time. Therefore, these results did not agree with original concept that the drinking of this decoction cause increases in urine flow as a diuretic. Thus, the increases in urine flow in that concept may be due to drinking a large amount of water. Besides, at highest concentrations, the decreasing of urine flow rate and urinary excretion of electrolytes and this may be due to hemoconcentrations to trigger the kidney to reabsorb water and electrolyte. However, the direct mechanism is still unclear and it needs to be searched for and investigated.

References


